



Review Article

Host-Pathogen Interactions in Immunocompromised States: A Comparative Study of Veterinary and HIV/AIDS Microbiology

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Received: 19 April 2026

Accepted: 27 April 2026

Published: 30 April 2026

DOI Prefix

10.59644

Quick Response Code:



ISSN (p): 2959-619X

ISSN (e): 2959-6203

Website: mdpi.com

Publisher: MDPIP

ABSTRACT

The purpose of this study is to compare the host–pathogen interactions in immunocompromised conditions, particularly human immunodeficiency virus/acquired immunodeficiency syndrome (HIV/AIDS) and veterinary lentiviral infections, such as feline immunodeficiency virus (FIV) and simian immunodeficiency virus (SIV). We adopted the Damage-Response Framework as the main paradigm to investigate immune vulnerabilities, evasion strategies of pathogens, clinical disease manifestation, and treatment approaches across species. Our review highlights a “mucosal catastrophe” with selective depletion of cluster of differentiation 4-positive T helper 17 and T helper 22 cells in gut-associated lymphoid tissue, resulting in gut barrier disruption, translocation of bacteria, and systemic chronic inflammation. Pathogens have sophisticated evasion strategies, including latent reservoir establishment, antigenic drift, restriction factor evasion, and immune checkpoint subversion. Zoonotic opportunistic pathogens (*Cryptosporidium* species, *Toxoplasma gondii*, *Mycobacterium avium*, *Rhodococcus equi*) are a serious concern for vulnerable human populations. The study is limited by the veterinary literature for non-lentiviral diseases and variable designs, which preclude quantitative meta-analysis. We conclude that veterinary models have been directly translated into human therapies, such as tenofovir and the feline immunodeficiency virus vaccine. Implications include integrated One Health surveillance networks, translational therapeutic studies, and the breakdown of academic silos between human and veterinary medicine to improve global public health and outcomes for immunocompromised hosts.

Keywords: Host-Pathogen Interactions, Immunocompromised States, HIV/AIDS, Feline Immunodeficiency Virus (FIV), One Health.

How to cite this article: Yameen, M., Saleem, R., Hussain, Z., Rehman, A., Ramzan, M., Tanveer, M., & Rehman, S. (2026). Host-Pathogen Interactions in Immunocompromised States: A Comparative Study of Veterinary and HIV/AIDS Microbiology. *Open Access Public Health and Health Administration Review*, 4(2), 29-44. [https://doi.org/10.59644/oaphhar.4\(2\).271](https://doi.org/10.59644/oaphhar.4(2).271)



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INTRODUCTION

Numerous biological factors help human immunodeficiency virus (HIV) to develop into acquired immunodeficiency syndrome (AIDS), and opportunistic infections (OIs) are some of the most important contributors to morbidity (Stump & Vande Woude, 2007). AIDS is clinically characterized by severe immunodeficiency, which causes an increased occurrence of these OIs and other malignancies. Nearly identical to this in veterinary medicine is the feline immunodeficiency virus (FIV), a lentivirus with properties similar to HIV that, in feline hosts, predisposes them to secondary infections and neurological diseases (Silva *et al.*, 2025). Moreover, the infection of simian immunodeficiency virus (SIV) in macaques is a leading model of human AIDS as it recapitulates the loss of CD4+ T cells and the resulting development of opportunistic diseases that are life-threatening (Kumar, 2025). Most of the OIs common among human HIV/AIDS patients are known to be natural zoonoses that vertebrates are known to transmit to humans either through direct contact, vectors, or contaminated food and water (Russotto, 2022).

The intricate interaction can be most effectively described by the Damage-Response Framework (DRF), which views microbial pathogenesis as the emergent consequence of interaction between a particular host and a particular microbe, rather than as a single characteristic of (Kimball *et al.*, 2026). The DRF underlines that clinical disease occurs only when the extent of host damage, be it by the pathogen, immune response by the host, or both, alters normal physiological homeostasis. It is necessary to consider the inclusion of veterinary microbiology into the investigation of human immunocompromised states because most of the emerging diseases have a zoonotic background and exhibit similar pathogenesis in other species (Chauhan *et al.*, 2025). The epidemiological information gained through veterinary study of animal-related infections by pathogens like *Toxoplasma gondii*, *Cryptosporidium*, and *Mycobacterium Bovis* is critical in controlling immunocompromised human populations. Finally, de-academicizing animal and human health will enable a one-disease framework to understand the changing epidemiology and microbiology of global infectious threats (Devi *et al.*, 2021).

This review aims to:

- Compare immunological responses of host-pathogen interactions in major veterinary immunocompromised states and HIV/AIDS.
- Determine typical opportunistic pathogens and their avoidance mechanisms.
- Test cross-species translational knowledge on therapeutic and preventive interventions.
- Suggest an integrated management approach to one-health.

METHODS AND MATERIALS

Search Strategy and Information Sources

This comparative review was prepared according to the recommendations for narrative and "systematic-like" reviews in microbiology and immunology. We conducted a very thorough search of electronic databases such as PubMed/MEDLINE, Scopus, Web of Science, and Google Scholar. The date range for the search was January 2000 to March 2026, to capture both seminal and recent lentiviral research. Furthermore, the bibliographies of obtained articles were searched for additional sources.

Search Terms and Keywords

The keywords used included, with Boolean operators (AND, OR): "host-pathogen interactions," "immunocompromised states," "HIV/AIDS," "feline immunodeficiency virus" OR "FIV," "simian immunodeficiency virus" OR "SIV," "opportunistic infections," "microbial translocation," "Th17 depletion," "mucosal catastrophe," "latent reservoir," "zoonotic pathogens," "One Health," and "veterinary lentivirus". Search terms were modified to suit each database using the controlled vocabulary (such as Medical Subject Headings for PubMed).

Inclusion and Exclusion Criteria

Inclusion criteria included: (i) original research, clinical trials, and peer-reviewed reviews; (ii) references to mechanisms of susceptibility (e.g., CD4+ T cell depletion, mucosal barrier dysfunction); (iii) descriptions of pathogen evasion strategies in human or veterinary lentiviruses; (iv) Information about clinical presentation, diagnostics, and treatments; and (v) English language

publications. Articles were excluded if they: (i) were not peer-reviewed, were editorials, or conference abstracts without data; (ii) reported on non-lentiviral immunocompromised conditions (e.g., chemotherapy-induced immunosuppression) without comparisons to HIV/FIV/SIV; and (iii) did not provide sufficient details of their methods.

Data Extraction and Synthesis

Data from selected articles were extracted by two independent reviewers (M.Y. and R.S.) on an extraction form. The information extracted was authors, publication year, study design, host species, pathogen type, immunological outcomes, evasion strategies, and clinical or therapeutic outcomes. Any discrepancies were discussed or referred to a third reviewer (M.R.) We undertook a qualitative synthesis using a comparative thematic approach, grouping information into five broad themes: immunological basis of susceptibility, evasion mechanisms, clinical presentation, therapeutic insights, and zoonotic One Health implications. The Damage-Response Framework was used as the conceptual framework to understand pathogenesis in different species.

THEORETICAL BUILDUP

Immunological Basis of Susceptibility in Immunocompromised Hosts

Host immunocompromised hosts are best considered in the Damage-Response Framework (DRF), according to which microbial pathogenesis is an emergent product of host-pathogen interactions but not a fixed microbial property. Disease in this framework is observed when physiological homeostasis is disturbed due to host damage as a result of a weak or over-strong immune response (Cribbs et al., 2020). The gradual loss of CD4+ T lymphocytes is the general hallmark of this vulnerability in not only human HIV/AIDS, but also the models of Feline Immunodeficiency Virus (FIV) and Simian Immunodeficiency Virus (SIV) (Teer et al., 2025). These cells are key orchestrators of the immune system; their loss in terms of both quantity and quality results in a severe deficiency in the ability to generate effective responses, both cellular and humoral, to opportunistic challenges (Mukherjee & Mishra, 2026).

Figure 1

Mechanism of Intestinal Barrier Breakdown

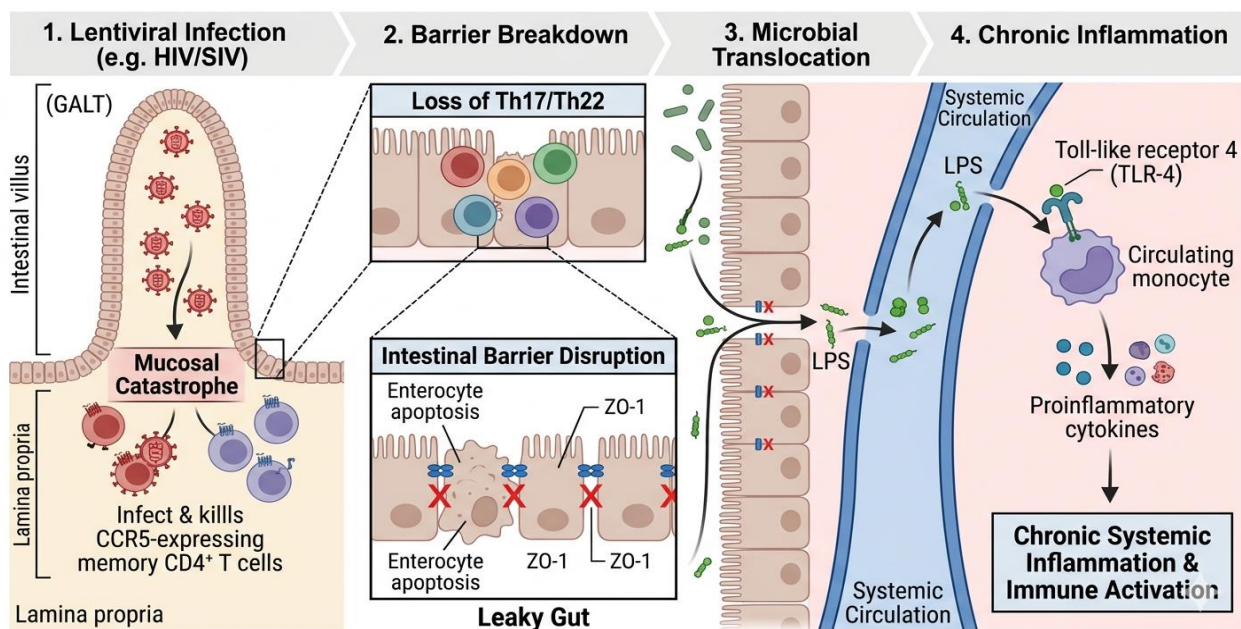


Figure 1: Sequential progression from lentiviral infection (e.g., HIV/SIV) in gut-associated lymphoid tissue (GALT) to chronic inflammation. (1) Selective infection of CCR5+ memory T cells by the virus (red dots). (2) Depletion of critical Th17/Th22 subsets disrupt epithelial homeostasis and tight junction proteins (ZO-1, marked with red 'X's). (3) The resulting leaky gut allows the systemic translocation of microbial products, including lipopolysaccharide (LPS). (4) Translocated LPS activates Toll-like receptor 4 (TLR-4) on circulating monocytes, driving chronic systemic immune activation and persistent inflammation, a stronger predictor of disease progression than viremia.

One of the key early events of this process is the "mucosal catastrophe" that takes place in the gut-associated lymphoid tissue (GALT) (Chen, 2023). The virus specifically infects and kills CCR5-expressing memory CD4+ T cells that are numerous in the intestinal mucosa. In particular, the depletion of Th17 and Th22 subsets is catastrophic because these cells play a crucial role in the preservation of the intestinal epithelial barrier and the generation of antimicrobial peptides. They are depleted and cause enterocyte apoptosis and tight junction disruption, causing a leaky gut (Alexandrova *et al.*, 2022). This anatomical defect promotes microbial translocation, in which intestinal luminal microbial products like lipopolysaccharide (LPS) are translocated into systemic circulation. LPS then causes Toll-like receptor 4 (TLR-4) and leads to chronic systemic immune activation and chronic inflammation, which are typically more reliable indicators of disease progression than viremia itself (Mu *et al.*, 2024). The mechanisms of cell death also make immunopathology complicated. Although direct viral lysis does take place, about 95 percent of CD4+ T cell loss in lymphoid tissues is caused by pyroptosis, which is a very inflammatory type of programmed cell death that is brought about by abortive infection (Lionakis *et al.*, 2023). This forms a vicious cycle that kills cells and releases proinflammatory cytokines, which attract and activate new target cells, creating new viral replication and tissue destruction. Moreover, the host experiences T cell exhaustion, a functional impairment caused by ongoing exposure to the antigen and characterized by an increase in inhibitory receptors such as PD-1 and CTLA-4 (Yang *et al.*, 2020). This fatigue restrains the proliferative ability of the effector cells, making the host incapable of eliminating the pathogen in a manner that a healthy system can deal with easily.

Moreover, Th1 to Th2 cytokine switch compromises cell-mediated immunity against intracellular infections, like Mycobacterium avium, and facilitates dysregulated B cell activation. Finally, the immunological pathogenesis is not an immunodeficiency but a complicated, dysregulated inflammation and immune senescence (Lv *et al.*, 2021). In order to supplement the discussion on the Immunological Basis of Susceptibility in Immunocompromised Hosts, the next table provides an overview of the key mechanisms observed in the sources that contribute to disease progression and susceptibility to opportunistic threats in HIV and veterinary lentiviral models.

Table 1

Key Immunological Mechanisms of Susceptibility in HIV and Veterinary Lentiviral Models (Chauvin & Sauce, 2022; Chen et al., 2023; Yan et al., 2023; Balendran et al., 2024)

| Mechanism | Description | Primary Markers/Cell Types Involved | Pathological Impact |
|-----------------------|---|-------------------------------------|---|
| CD4+ T Cell Depletion | Quantitative and qualitative loss of central immune coordinators. | CD4+ T lymphocytes | Hallmark of immunodeficiency: increased OIs and malignancy. |
| Mucosal Catastrophe | Rapid elimination of memory T cells in the gut during primary infection. | CCR5+ memory CD4+ T cells | Early structural damage to the gut-associated lymphoid tissue (GALT). |
| Th17 and Th22 Loss | Depletion of subsets is vital for maintaining mucosal integrity and antimicrobial peptides. | Th17 and Th22 cells; IL-17, IL-22 | Impaired barrier repair and loss of mucosal antimicrobial defense. |

| | | | |
|-------------------------------|---|--|--|
| Intestinal Barrier Disruption | Disruption of epithelial tight junctions and enterocyte apoptosis. | ZO-1 tight junction protein | Development of "leaky gut," allowing systemic pathogen entry. |
| Microbial Translocation | Systemic escape of microbial products (e.g., LPS) from the intestinal lumen. | Lipopolysaccharide (LPS); sCD14 | Chronic systemic immune activation and persistent inflammation. |
| T Cell Exhaustion | State of functional impairment due to persistent antigen exposure. | PD-1, CTLA-4, Tim-3 | Reduced proliferative capacity and failure to clear viral/opportunistic threats. |
| Cytokine Shift | Imbalance favoring regulatory or humoral responses over cell-mediated immunity. | Th1 to Th2 shift; TGF- β , IL-10 | Impaired response against intracellular pathogens like <i>M. avium</i> . |
| Pyroptosis | Highly inflammatory programmed cell death triggered by abortive infection. | Caspase-1; IL-1 β | Vicious cycle of inflammatory cell recruitment and tissue damage. |

This immunological environment depicts that susceptibility is not simply a deficiency of immune response, but of chronic, dysregulated inflammation and structural breakdown at the mucosal surfaces. Inability to replace these particular mucosal subsets together with the functions of barriers despite suppressive antiretroviral treatment has been one of the most pressing issues in clinical treatment.

Pathogen Evasion Strategies across Host Species

To overcome host defense, pathogens have developed a rich repertoire of strategies, and this is most productively described in the Damage-Response Framework (DRF), where the pathogenesis is an emergent property of host-microbe interactions instead of a fixed microbial characteristic (Obeagu & Obeagu, 2024). One of the main mechanisms common to human and veterinary lentiviruses is the creation of latent reservoirs, most frequently in the resting memory CD4+ T lymphocytes and lymphoid tissues. In human HIV-1 and feline immunodeficiency virus (FIV), transcriptionally silent proviruses are integrated into the host genome, and this enables them to be undetectable by cytolytic effectors and forms a permanent shield against elimination (Gunawan *et al.*, 2026). Moreover, these viruses take advantage of anatomic niches like the central nervous system (CNS), the gastrointestinal tract (GIT), and B cell follicles, wherein ineffective drug penetration and limited immune surveillance allow autonomous viral evolution and survival.

Antigenic variation is an important mechanism of escape, especially for viruses with high mutation rates. HIV-1 constantly produces the so-called decoy proteins and viral mutations in order to counteract the earlier antibody and cytotoxic T lymphocyte (CTL) responses (Ding *et al.*, 2026). Equally, equine infectious anemia virus (EIAV) employs sequence variation in its Rev protein to limit the expression of structural genes at times of clinical quiescence, thus causing an evasion of the host immune system (Su *et al.*, 2022). In addition to viruses, *Cryptosporidium* species have a great deal of intraspecific variation in their sub-telomeric genes, such as the MEDLE, insulinase-like proteins (INS), and SKSR families. These putative virulence factors genomic islands enable the parasite to evolve into host species and tune immune responses (Bashiardes *et al.*, 2026).

Pathogens also synthesize certain proteins to resist host restriction factors, which are proteins in the innate cell that are meant to prevent viral replication. HIV-1 Vif protein causes the degradation of human APOBEC3G to avoid disastrous G-to-A

hypermutations in the viral genome (He *et al.*, 2025). HIV-1 and HIV-2 also use Vpu and Env to antagonize tetherin, which is a protein that inhibits the release of nascent virions of the host cell, respectively. In veterinary systems, the simian immunodeficiency virus (SIV) inverts SAMHD1 through the expression of a protein called Vpx, which is an accessory protein that allows the virus to infect non-dividing macrophages and dendritic cells; a process that is absent in HIV-1 (Jasinska *et al.*, 2022). Also, HIV and SIV viral Nef proteins selectively suppress MHC-I molecules to prevent the expression of viral epitopes to the adaptive immune system (Aldhalmi & Al-hadrawi, 2022).

Intracellular opportunistic pathogens use structural and behavioral adaptations to survive in immunocompromised conditions. According to the Amoeboid Predator-Animal Virulence Hypothesis, such pathogens as *Mycobacterium avium* and *Legionella pneumophila* use environmental amoeba as a bootcamp to develop resistance to mammalian macrophages (Navasardyan *et al.*, 2023). Such pre-adaptation enables them to use host phagocytes as protective niches.

Talaromyces marneffei employs thermal dimorphism to convert a mold into a pathological phase of a yeast, which enables it to endure endothermic hosts. It also alters the immune microenvironment through M2 macrophage polarization and programmed death-ligand 1 (PD-L1) upregulation, which basically transforms host cells into Trojan horses to be disseminated throughout the system without being detected by the immune system (Augello *et al.*, 2023). Similarly, *Cryptosporidium* sporozoites develop a secure niche at the host epithelial apex, creating a parasitophorous vacuole and a specialized feeder organelle to scavenge nutrients but are not a part of the host cytosol. Lastly, the capability of pathogens such as *Cryptosporidium* to resist the environment is augmented by the formation of thick-walled oocysts, which are highly resistant to chemical and physical adversities (Obeagu & Obeagu, 2026).

Table 2

Comparative Pathogen Evasion Strategies in Human and Veterinary Microbiology (Scaglioni, 2025; Le Hingrat et al., 2021; Wang et al., 2021; Jiménez et al., 2023; Tokarev et al., 2022)

| Evasion Strategy | Mechanism | Examples (Human/Veterinary) |
|-----------------------------------|---|---|
| Latent Reservoirs | Integration into the host genome in resting memory cells. | HIV-1, FIV, SIV (Resting memory CD4+ T cells) |
| Anatomic Sanctuaries | Persistence in sites with poor drug penetration (CNS, GALT). | HIV-1, FIV, SIV |
| Antigenic Variation | Rapid mutation of surface glycoproteins and regulatory proteins. | HIV-1 (Env), EIAV (Rev), <i>Cryptosporidium</i> (gp60) |
| Restriction Factor Counteraction | Degradation or antagonism of innate cellular blocks. | HIV-1 (Vif vs APOBEC3G; Vpu vs Tetherin); SIV (Vpx vs SAMHD1) |
| Antigen Presentation Interference | Downregulation of MHC-I to avoid CTL detection. | HIV-1 (Nef), Veterinary Lentiviruses |
| Host Cell Polarization | Induction of M2 macrophage phenotype for safe residence. | <i>Talaromyces marneffei</i> , <i>Toxoplasma gondii</i> , <i>M. avium</i> |
| Immune Checkpoint Manipulation | Upregulation of inhibitory ligands like PD-L1. | <i>Talaromyces marneffei</i> |
| Structural Protection | Formation of parasitophorous vacuoles or thick-walled oocysts. | <i>Cryptosporidium</i> spp., <i>Microsporidia</i> |
| Thermal Dimorphism | Phase transition (mold to yeast) to survive at the host body temperature. | <i>Talaromyces marneffei</i> , <i>Histoplasma</i> spp. |

Figure 2
Comparative Immune Evasion by Human and Veterinary Pathogens

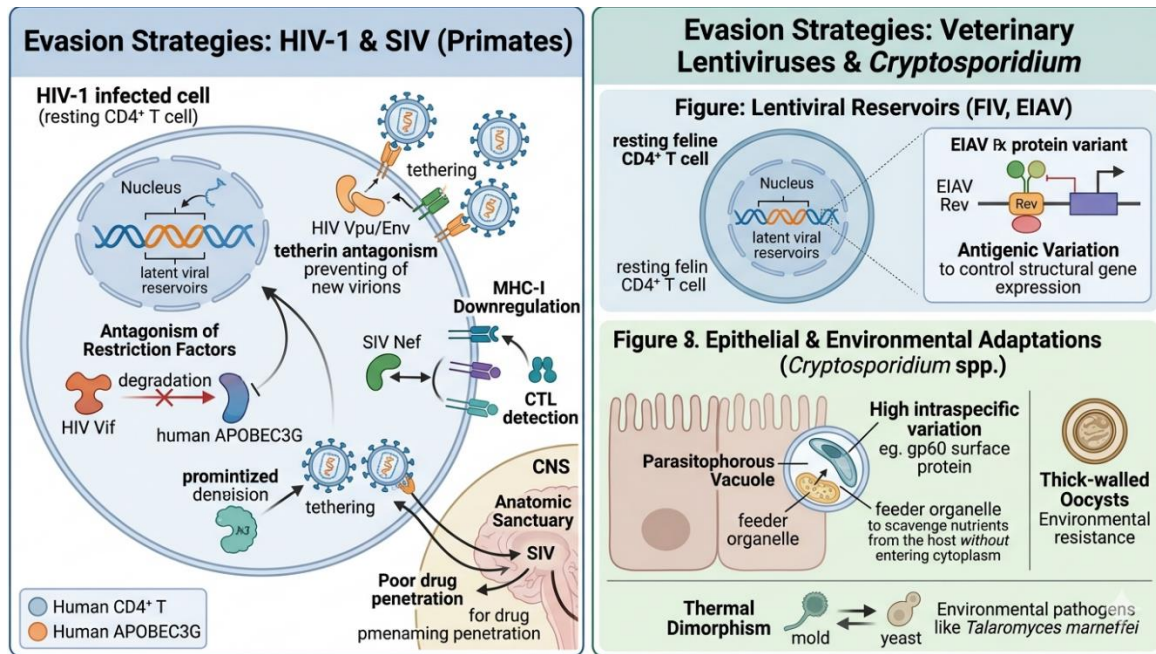


Figure 2: This visual summary illustrates analogies in microbial pathogenesis across species. (Left Panel) Mechanisms observed in HIV-1 and SIV infection, including the establishment of latent viral reservoirs in the nucleus, antagonism of restriction factors (Vif degrading APOBEC3G; Vpu/Env antagonizing tetherin), downregulation of MHC-I to evade CTL detection (Nef), and the use of the CNS as an anatomic sanctuary. (Right Panel) Evasion in veterinary lentiviruses and zoonotic enteric pathogens. Similar reservoirs are formed by FIV; EIAV exhibits high antigenic variation (Rev protein) to control structural gene expression. The diagram of *Cryptosporidium* shows unique adaptations: developing a distinct parasitophorous vacuole and feeder organelle at the epithelial apex (blue arrows) to survive extracellularly from the cytoplasm, alongside environmental resistance (thick-walled oocysts). A shared mechanism of thermal dimorphism is also visualized for environmental pathogens like *Talaromyces marneffei*.

Clinical Manifestations and Diagnostic Approaches

Diseases in immunocompromized states are mainly mediated by the progressive loss of CD4+ T lymphocytes, which is the main surrogate endpoint in terms of monitoring disease severity and predicting the overt development of immunodeficiency in humans and veterinary animals. In human HIV infection, this loss results in a shift after an acute phase (which is usually marked by anorexia and lymphadenopathy) into clinical AIDS, which is marked by a CD4 count below 200/mm³ and the development of opportunistic infections (OIs) and malignancies at high incidence (Okoye & Picker, 2013). This pathology is also well reflected in veterinary medicine by Feline Immunodeficiency Virus (FIV), which has a long period of asymptomatic infection and a symptomatic period with periodic relapsing secondary infections, wasting, and B-cell or T-cell lymphomas (McCune, 2001). Respiratory signs and symptoms constitute a major burden of morbidity in the world. *Mycobacterium tuberculosis* is the most prevalent significant OI in humans, and it tends to hasten the development of HIV and elevate the rate of mortality. This is similar in veterinary medicine to *Rhodococcus equi* in horses that causes cavitary, granulomatous pneumonia resembling human tuberculosis (Février *et al.*, 2011). Moreover, *Pneumocystis jiroveci* (PJP) is also a hallmark of human AIDS and FIV-infected cats, and is the major cause of severe pneumonia, which is caused by the destruction of alveolar spaces. Fungal coinfections such as *Aspergillus* have become significant hazards and commonly complicate cases of previously treated pulmonary tuberculosis by colonizing and forming aspergillomas in the remaining lung cavities (Alimonti *et al.*, 2003).

Gastrointestinal involvement is a cross-species finding and is usually associated with severe watery diarrhea and dehydration. *E. coli*, *Cryptosporidium* species, and other pathogens result in the major destruction of epithelial tight junctions and the loss of barrier function, and the inability to absorb nutrients. This "mucosal catastrophe" promotes microbial translocation, in which microbial products like lipopolysaccharide (LPS) are released outside the intestinal lumen into the systemic circulation and cause chronic inflammation in both humans and bovine hosts (Vidya Vijayan *et al.*, 2017). Salmonella bacteremia that occurs regularly can be identified as an AIDS-defining diagnosis, which indicates the presence of severe underlying immune malfunction. On the same note, *Campylobacter* Jejuni has been commonly implicated in both human and animal diarrhea, pointing to the commonality in risk factors that contribute to the zoonotic enteric threats (Peng *et al.*, 2020). Additional neurological and dermatological manifestations demonstrate the breadth of clinical disease. Neurotropic strains of HIV and FIV infect CNS macrophages, microglial cells, and astrocytes, causing HIV-associated neurocognitive disorders (HAND) in humans or feline neurological abnormalities, including aggressive behavior and delayed reflexes (Brenchley *et al.*, 2004). A perfect example of a zoonotic threat is *Toxoplasma gondii*; cats are the host organisms, oocysts of which can cause deadly encephalitis to humans with weakened immunity. These clinical features are usually accompanied by chronic inflammatory diseases such as stomatitis and gingivitis in cats, which tend to respond to treatment with zidovudine (Beck, 2005).

Diagnostic methods should be multi-modal in distinguishing between active clinical disease and simple infection. Quantitative PCR of viral load and CD4 flow cytometry of CD4 counts are still the gold standard for human HIV. ELISA and Western blot are widely used in veterinary settings to detect lentiviruses, but the best cost and infrastructure frequently restrict access to nested PCR (Tolomeo & Cascio, 2024). In the case of human herpesviruses (HHVs) that are important copathogens, laboratory techniques encompass antigen tests in tumor tissue biopsies as well as RT-PCR of body fluids. Recent developments imply the use of soluble CD14 (sCD14) and I-FABP as biomarkers to determine monocyte activation and enterocyte injury as a result of microbial translocation. Moreover, pathogen screening, such as cryptococcal antigen (CrAg) and interferon-gamma release assays (IGRA), should be used in the initial stages of screening in high-risk groups (Mishra *et al.*, 2009). Finally, the key to successful management is to promptly detect these coinfections and avoid the development of immune reconstitution inflammatory syndrome (IRIS) upon the initiation of treatment.

Therapeutic and Preventive Strategies: A Comparative View

The treatment of immunocompromised conditions has undergone a paradigm shift, and diseases such as HIV/AIDS are no longer terminal diagnoses but a manageable chronic illness with the amazing success of Highly Active Antiretroviral Therapy (HAART). This medical advancement finds close reflection in veterinary medicine, in which human antiretroviral drugs have been repackaged to treat lentiviral infections of cats and monkeys, and is an example of a continuity of pharmacological development that can be termed as One Health (Foka & Mufhandu, 2023). The standard of care in humans is a combination regimen, usually two nucleoside reverse transcriptase inhibitors (NRTIs) with an integrase inhibitor or a protease inhibitor, to maintain viral control. This is not only a way to decrease morbidity but also a Treatment as Prevention (TasP), as it is known to decrease the risk of transmission as the infectiousness of the host is minimized (Bakasis & Androutsakos, 2021). Likewise, in feline medicine, zidovudine (AZT) is effective in enhancing the clinical condition of cats with FIV, especially in the treatment of related stomatitis as well as in the correction of CD4/CD8 ratios. Non-species toxicity of the drugs, however, tends to limit their use in veterinary practice, including non-regenerative anemia in cats receiving adefovir (PMEA) or bone marrow suppression with high-dose AZT. Interestingly, the design of human therapies based on Tenofovir was actually influenced by preliminary studies of efficacy in FIV-infected cats (Alemu *et al.*, 2022).

Preventative measures are another area of cross-species synergy that is also of major concern, especially in the field of vaccinology. As an efficient human HIV-1 vaccine has not been developed even after decades of intensive research and promising trials, such as RV144, veterinary medicine has made a breakthrough with the commercial launch of a dual-subtype FIV vaccine (Fel-O-Vax FIV) (Kamvuma *et al.*, 2026). This inactivated whole-virus vaccine has a global subtype and circulating recombinant prophylaxis, and its efficacy has mainly been credited to the development of strong T-cell immunity. The veterinary

accomplishments offer crucial models to human vaccine development, especially in the discovery of evolutionarily preserved epitopes on proteins such as p24 that may induce cross-protective cellular responses. In addition to vaccines, preventive care includes vigorous screening and prophylaxis against opportunistic infections (OIs) (Sun & Wang, 2024). An example is the use of acyclovir to slow the disease course in HIV/HSV-2 coinfecting persons, and the use of trimethoprim-sulfamethoxazole as the basis of prophylaxis against *Pneumocystis jirovecii* pneumonia.

The contemporary treatment edge has now shifted to destroying latent reservoirs and alleviating chronic inflammation. In human studies, the goal of the "Shock and Kill" approach is to activate the expression of viruses in resting memory CD4⁺ T cells by administration of latency-reversing agents, namely HDAC inhibitors, and induce their immune clearance. Preclinical testing of these interventions would not be possible without veterinary models, especially the SIV-infected macaque (Miti et al., 2020). Moreover, since ART does not fully heal the mucosal state, researchers are exploring adjunct interventions to treat microbial translocation. Administration of the non-systemic antibiotic rifaximin or probiotics has demonstrated the potential of reducing systemic immune activation in SIV-infected macaques and in HIV-infected humans. Lastly, zoonotic risk reduction should be incorporated in clinical management. In the case of immunocompromised people, this involves observing stringent hygiene protocols when contacting pets, including not feeding animals raw meat and the use of gloves during litter box cleaning to avoid contacting pathogens like *Toxoplasma gondii* and *Salmonella* (Saka et al., 2023). Treatment then requires a concerted effort between doctors and veterinarians to treat the common microbial habitat of the host.

Zoonotic Dynamics and the One Health Perspective in Immunocompromised Populations

The syndemic model of the clinical manifestation of infectious disease in immunocompromised states is becoming more common, in which the potential of a pathogen is mediated by the interaction between it and other infectious agents and environmental factors. Many life-threatening opportunistic infections (OIs) in human HIV/AIDS patients are known as zoonoses, which are naturally acquired between human beings and vertebrate animals (Ullah, 2026). This fact requires dismantling of academic isolations between human and animal health, and the implementation of the so-called one disease model that has long been a mainstay of veterinary medicine to safeguard human health around the world (Hossain et al., 2025).

Rhodococcus equi, a common veterinary pathogen in horses, is one of the most important bacterial zoonoses, causing cavitary granulomatous pneumonia in immunocompromised humans, which is similar in appearance to pulmonary tuberculosis. Likewise, nontyphoidal *Salmonella* and *Campylobacter jejuni* are common isolates of both human and animal enteric infections, and repeated *Salmonella* bacteremia is a definitive AIDS-defining diagnosis (Wilcox & Steele, 2021). The importance of domestic animals as reservoirs is also found in *Bartonella* species, including *B. henselae* (the cause of cat-scratch disease), which may induce bacillary angiomatosis and peliosis hepatis in patients with low CD4 counts. Moreover, protozoal parasites such as *Cryptosporidium* species have a high genetic diversity and a large host range, frequently having a livestock origin that results in disastrous mucosal catastrophes in humans with long-term systemic inflammation and microbial translocation (Hossain et al., 2025).

It is essential to integrate veterinary microbiology into clinical management of humans since studies of animal-related infections with such pathogens as *Toxoplasma gondii* and *Mycobacterium avium* offer valuable epidemiological data. Also, the ecological study of environmental pathogens such as *Legionella pneumophila* proposes an "Amoeboid Predator-Animal Virulence Hypothesis" in which microbes utilize environmental amoeba to pre-adapt to survive in mammalian macrophages. Finally, the immunocompromised need to be informed on how to reduce the risk of zoonotic infections by providing specific hygiene information about handling pets and livestock (Vale, 2022).

Table 3

Comparative Zoonotic Pathogens and Their Veterinary/Human Impact (Ellwanger et al., 2021; Ortiz-Millán, 2025; Overgaauw et al., 2020; Erkyihun & Alemayehu, 2022; Rauf et al., 2023)

| Pathogen | Primary Animal Reservoir | Veterinary Manifestation | Human Manifestation in Immunocompromised |
|----------------------|--------------------------|------------------------------|---|
| Rhodococcus equi | Horses (foals) | Granulomatous pneumonia | Cavitary pneumonia; TB-like disease |
| Bartonella henselae | Cats | Usually, asymptomatic | Bacillary angiomatosis; peliosis hepatis |
| Cryptosporidium spp. | Livestock, pets | Scours (watery diarrhea) | Wasting; mucosal catastrophe; "leaky gut" |
| Toxoplasma gondii | Felines (felids) | Asymptomatic oocyst shedding | Encephalitis; cerebral mass lesions |
| Mycobacterium avium | Birds, environment | Granulomatous disease | Disseminated infection; Th2 cytokine shift |
| Salmonella spp. | Livestock, reptiles | Enteritis; septicemia | Recurrent bacteremia; AIDS-defining illness |

DISCUSSION

Comparative study of human and veterinary microbiology demonstrates that states of immunocompromised have a common basic pathophysiological structure. Pathogenesis, in the context of the Damage-Response Framework (DRF), is an emergent consequence of the unique interaction of a host and a microbe and not a fixed microbial trait. This paradigm shift shifts the emphasis away, however, to reductionist "virulence factors" to the dynamic imbalance of host homeostasis, in which disease is expressed, either as a consequence of inadequate or excessive immune response (Dunga *et al.*, 2025).

One of the key themes that unite all the species is the mucosal catastrophe that occurs early during the infection. The intestinal epithelial barrier is destroyed by the selective depletion of Th17 and Th22 CD4 + T cell subsets in the gut-associated lymphoid tissue (GALT) (Samadi & Hailat, 2021). This structural breakdown results in microbial translocation- the systemic release of microbial products such as lipopolysaccharide (LPS)- a major cause of chronic systemic immune response and chronic inflammation in human HIV/AIDS and veterinary models such as Simian Immunodeficiency Virus (SIV) (Kundu *et al.*, 2024).

The essential templates of study of these complex interactions are done using veterinary models, especially Feline Immunodeficiency Virus (FIV) in cats and SIV in macaques. SIV infection in Asian macaques accurately recapitulates the characteristic CD4+ T cell depletion and consequent development of life-threatening opportunistic infections (OIs) and malignancies seen in human AIDS. In addition, a number of these OIs are known zoonoses like Cryptosporidium, Toxoplasma gondii, and Mycobacterium Bovis, which are naturally spread between vertebrate animals and humans (Gado *et al.*, 2023). The high rates of these infections, particularly in the developing regions, highlight the importance of the One Health approach that helps in communication between physicians and veterinarians in order to safeguard the health of the entire world. Veterinary research has had a direct clinical influence on human clinical practice, therapeutically. An example is that the original in vivo efficacy of acyclic nucleoside phosphonate analogues in FIV-infected cats was the basis of the development of Tenofovir, now a key component of human HIV treatment. Moreover, the commercial availability of the dual-subtype FIV vaccine (Fel-O-Vax FIV) presents a crucial template to human vaccine development, showing that extensive prophylactic efficacy could be attained by induction of strong, evolutionarily preserved T-cell immunity (Mubareka *et al.*, 2023).

With HIV/AIDS becoming a chronic disease extending over multiple decades, clinical attention has shifted away from acute OIs and towards persistent inflammation and non-AIDS comorbidities that are related to premature aging. The elimination of latent reservoirs and restoration of mucosal barriers should be a priority in future research (Hartady *et al.*, 2026). New approaches, like mitochondrial rescue through PPAR α signaling and probiotics (such as *Lactobacillus plantarum*), have the potential to heal the intestinal integrity regardless of immune cell regeneration. Finally, it is necessary to disrupt academic barriers to support comparative immunology to gain insight into the changing microbiology of immunocompromised hosts.

CONCLUSION

Immunocompromised states of host-pathogen interactions demonstrate an impressive biological continuum between human HIV/AIDS and veterinary lentiviral infections, especially Feline Immunodeficiency Virus. By utilizing the Damage-Response Framework (DRF), the paradigm shift of reductionist perceptions can be made, with pathogenesis being considered as an emergent, dynamic product of host-microbe interactions in which clinical disease only occurs when damage to homeostasis and physiological processes occurs. One key similarity of species is the early mucosal disaster, in which Th17 and Th22 subsets of CD4+ T cells are selectively depleted to cause intestinal barrier breakdown and microbial translocation. This structural breakdown promotes chronic systemic immune activation, defining human and veterinary lentiviral disease. Translational antiretroviral development, especially with feline and simian models, will continue to rely on veterinary research to develop antiretrovirals such as Tenofovir and develop effective prophylaxis vaccines. Moreover, the high rates of life-threatening zoonotic opportunistic infections, including *Cryptosporidium* and *Toxoplasma gondii*, highlight the urgent need for a One Health model, which will help to communicate across sectors. Finally, the shift of HIV/AIDS into a chronic illness necessitates the development of therapeutic measures in the future beyond viral suppression to eliminate latent reservoirs and develop novel repair of mucosal barriers.

LIMITATIONS

There are several limitations to this review. First, the literature on veterinary immunocompromised states (especially BIV and non-lentiviral conditions) is limited in comparison to HIV/AIDS. Second, study designs (case reports versus cohort studies) are heterogeneous, which does not allow direct quantitative comparisons. Third, the majority of veterinary literature does not include immune phenotyping (e.g., CD4:CD8 ratios) that can be standardized to allow meta-analysis. Lastly, the review lacked non-mammalian immunocompromised models (e.g., avian), which can provide further information.

Limitations of the Review Methodology

Meta-analysis was unavailable because of different study designs, species-specific outcome measures, and a lack of consistency in immunophenotyping (e.g., CD4:CD8 ratio in veterinary studies). We did not include grey literature or non-English sources, potentially leading to a language bias.

RECOMMENDATIONS

It is based on this comparative review that the following recommendations are made to improve the clinical management and scientific understanding of immunocompromised hosts:

Integrated Surveillance Networks: Develop cross-sectoral surveillance of opportunistic infections in people and animals, to support a one-disease model to safeguard the world population's health. It is important to monitor the occurrence and distribution of pathogens like *Rhodococcus equi*, *Mycobacterium Bovis*, and *Salmonella* as these are major zoonotic agents to HIV-infected people.

Cross-Species Therapeutic Trials: Test repurposed antiviral and immunomodulatory agents in FIV-positive cats to guide human HIV adjunctive therapies. The effectiveness of feline models to showcase the effectiveness of acyclic nucleoside phosphonates, which was the direct basis of the development of Tenofovir, exemplifies the translational importance of these comparative trials.

Future studies should also focus on immunomodulators with the ability to suppress chronic systemic inflammation, a common hallmark of lentiviral disease.

Diagnostic Accessibility: Work on low-cost point-of-care (POC) PCR and CD4 count diagnostic tests in veterinary clinics, especially in low-resource environments. Use of sensitive diagnostics should be standardized in veterinary medicine because a lot of states associated with animals have immunocompromised states that are currently not researched rigorously enough to allow comparative research.

Public Health Education: Educate people with weakened immunity on zoonotic risk of pets and livestock, and focus on preventive behaviors like hand hygiene, no diets of raw meat in animals, and the use of gloves when cleaning litter boxes. Better interaction between physicians and veterinarians is essential because, in a clinical setting, there is a lack of accurate information on the risks of animal-human bonding.

Comparative Research Funding: Provide specific research funding for comparative immunology and One Health infectious disease programs. The bridging of human and veterinary microbiology longitudinal cohort studies is needed to determine evolutionarily conserved immune targets and to comprehend the intricacies of host-pathogen interactions in immunological failure states.

DECLARATION

Ethical Consideration: This study strictly adhered to the Declaration of Helsinki and relevant national and institutional ethical guidelines. Informed consent was obtained. All procedures performed in this study were consistent with the ethical standards of the Declaration of Helsinki. This review article does not involve primary data collection from human or animal subjects. All cited studies adhered to their respective institutional ethical guidelines.

Conflict of Interest: The authors have no conflict of interest.

Consent for Publication: The authors give consent for publication.

Availability of Data and Materials: All data analyzed in this review are from published articles cited in the references. No primary data were generated. Data could be provided subject to a written request from the corresponding author.

Funding Source: Not applicable.

Acknowledgement: The authors gratefully acknowledge all people for their valuable support and help.

Use of Artificial Intelligence (AI)-Assisted Technology for Manuscript Preparation: Artificial intelligence (AI) tools were used solely to assist with language editing. No AI tools were used for data extraction, statistical analysis, result interpretation, or the generation of original scientific content. All analyses were conducted by the authors, and they take full responsibility for the integrity and accuracy of the manuscript.

Similarity Index/ Plagiarism: The similarity index was checked, and it is well below the threshold value of 19%, whereas each source is less > 5%.

Authors' Contributions: All the authors contributed equally.

REFERENCES

- Aldhalmi, H. K., & Al-hadrawi, K. K. (2022). Immunopathogenesis of HIV/AIDS: Linking Viral Replication, Immune Dysregulation, and Systemic Patient Challenges. *Organization*, 30, 51.
- Alemu, G. G., Nigusie, Z. M., Amlak, B. T., & Achamyeleh, A. A. (2022). Survival time and predictors of death among HIV-infected children under five after initiation of anti-retroviral therapy in West Amhara Referral Hospitals, Northwest Ethiopia. *BMC Pediatrics*, 22(1), 670.
- Alexandrova, Y., Costiniuk, C. T., & Jenabian, M. A. (2022). Pulmonary immune dysregulation and viral persistence during HIV infection. *Frontiers in Immunology*, 12, 808722.


- Alimonti, J. B., Ball, T. B., & Fowke, K. R. (2003). Mechanisms of CD4+ T lymphocyte cell death in human immunodeficiency virus infection and AIDS. *Journal of General Virology*, 84(7), 1649-1661.
- Augello, M., Bono, V., Rovito, R., Tincati, C., & Marchetti, G. (2023). Immunologic interplay between HIV/AIDS and COVID-19: adding fuel to the flames? *Current HIV/AIDS Reports*, 20(2), 51-75.
- Bakasis, A. D., & Androutsakos, T. (2021). Liver fibrosis during antiretroviral treatment in HIV-infected individuals. Truth or tale? *Cells*, 10(5), 1212.
- Balendran, T., Iddawela, D., & Lenadora, S. (2024). Cryptosporidiosis in a zoonotic gastrointestinal disorder perspective: present status, risk factors, pathophysiology, and treatment, particularly in immunocompromised patients. *Journal of Tropical Medicine*, 2024(1), 6439375.
- Bashiardes, S., Heinemann, M., Adlung, L., Valdés-Mas, R., Mahdi, J. A., Nobs, S. P., & Elinav, E. (2026). Human immunodeficiency virus-associated gut microbiome impacts systemic immunodeficiency and susceptibility to opportunistic gut infection. *Nature Microbiology*, 1-14.
- Beck, J. M. (2005). *The immunocompromised host: HIV infection*. Proceedings of the American Thoracic Society, 2(5), 423-427.
- Brenchley, J. M., Schacker, T. W., Ruff, L. E., Price, D. A., Taylor, J. H., Beilman, G. J., ... & Douek, D. C. (2004). CD4+ T cell depletion during all stages of HIV disease occurs predominantly in the gastrointestinal tract. *The Journal of Experimental Medicine*, 200(6), 749-759.
- Chauhan, M., Shivarathri, R., Aptekmann, A. A., Chowdhary, A., Kuchler, K., Desai, J. V., & Chauhan, N. (2025). The Gcn5 lysine acetyltransferase mediates cell wall remodeling, antifungal drug resistance, and virulence of *Candida auris*. *MSphere*, 10(4), e00069-25.
- Chauvin, M., & Sauce, D. (2022). Mechanisms of immune aging in HIV. *Clinical Science*, 136(1), 61-80.
- Chen, J., Shao, J., Dai, M., Fang, W., & Yang, Y. L. (2023). Adaptive immunology of *Cryptococcus neoformans* infections – an update. *Frontiers in Immunology*, 14, 1174967.
- Cribbs, S. K., Crothers, K., & Morris, A. (2020). Pathogenesis of HIV-related lung disease: immunity, infection, and inflammation. *Physiological Reviews*, 100(2), 603-632.
- Devi, P., Khan, A., Chattopadhyay, P., Mehta, P., Sahni, S., Sharma, S., & Pandey, R. (2021). Co-infections as modulators of disease outcome: minor players or major players? *Frontiers in Microbiology*, 12, 664386.
- Ding, X., Fan, L., Xu, L., Ma, X., Meng, P., Li, J., ... & Yue, J. (2024). Incomplete immune reconstitution and Traditional Chinese Medicine in patients with HIV/AIDS: challenges and perspectives. *Infection and Drug Resistance*, 5827-5838.
- Dunga, K. E., Okoro, C. I., Onyenama, A. C., Ekuma, U. O., Ohanusi, I. N., & Izah, S. C. (2025). Implementing One Health approach to emerging zoonotic diseases: Bridging surveillance, sustainability, and global governance. *Exon*, 2(3), 200-223.
- Ellwanger, J. H., Veiga, A. B. G. D., Kaminski, V. D. L., Valverde-Villegas, J. M., Freitas, A. W. Q. D., & Chies, J. A. B. (2021). Control and prevention of infectious diseases from a One Health perspective. *Genetics and Molecular Biology*, 44(1 Suppl 1), e20200256.
- Erkyihun, G. A., & Alemayehu, M. B. (2022). One Health approach for the control of zoonotic diseases. *Zoonoses*, 2(1), 963.
- Fevrier, M., Dorgham, K., & Rebollo, A. (2011). CD4+ T cell depletion in human immunodeficiency virus (HIV) infection: role of apoptosis. *Viruses*, 3(5), 586.
- Foka, F. E. T., & Mufhandu, H. T. (2023). Current ARTs, virologic failure, and implications for AIDS management: a systematic review. *Viruses*, 15(8), 1732.
- Gado, D. A., Ehizibolo, D. O., Meseko, C. A., Anderson, N. E., & Lurz, P. W. (2023). Review of emerging and re-emerging zoonotic pathogens of dogs in Nigeria: Missing link in one health approach. *Zoonotic Diseases*, 3(2), 134-161.
- Gunawan, C. K., Sumarpo, A., & Indrati, A. R. (2026). Immune Dysregulation in HIV-TB Co-Infection: Role of Cytokines and T Cell Biomarkers-A Narrative Review. *Pathogens*, 15(1), 51.
- Hartady, T., Satrio, F. A., Maulana, S., Wira, D. W., Setyowati, E. Y., & Salleh, A. (2026). Zoonotic Tuberculosis as a One Health Challenge: Global Evidence, Transmission Dynamics, and Policy Gaps in Indonesia. *Veterinary Sciences*, 13(3), 237.
- He, J., Tan, S., & Lv, J. (2025). Tripartite interplay: immune reconstitution dynamics in AIDS, gut microbiota, and *Helicobacter pylori* infection: current advances and therapeutic prospects. *Gut Pathogens*, 17(1), 50.

- Hossain, D., Saeed, S. I., Ajose, D. J., Egbu, C. F., Adesola, R. O., Ogundijo, O. A., ... & Bristi, S. Z. T. (2025). Global Zoonotic Diseases and Public Health: A One Health Perspective. *One Health Integration: Global Perspectives on Animal Health and Sustainable Agriculture*, 165-208.
- Hossain, H., Chowdhury, M. S. R., Khan, S. S., Ahmad, T., Brishty, K. A., Rahman, M., ... & Rahman, M. M. (2025). Emerging Zoonotic Diseases: Epidemiology, Public Health Impact, and the Urgent Need for a Unified "One Health" Approach. *Pakistan Veterinary Journal*, 45(1).
- Jasinska, A. J., Pandrea, I., & Apetrei, C. (2022). CCR5 as a coreceptor for human immunodeficiency virus and simian immunodeficiency viruses: a prototypic love-hate affair. *Frontiers in Immunology*, 13, 835994.
- Jiménez, V. C., Geretz, A., Tokarev, A., Ehrenberg, P. K., Deletsu, S., Machmach, K., ... & Bolton, D. L. (2023). AP-1/c-Fos supports SIV and HIV-1 latency in CD4 T cells infected in vivo. *Iscience*, 26(10).
- Kamvuma, K., Hamooya, B. M., Mulemena, J. A., Masenga, S. K., Kirabo, A., & Munsaka, S. M. (2026). Incidence and correlates of chronic anemia in people living with HIV Initiating ART: An exploratory five-year retrospective cohort of patients retained in care. *PLOS Global Public Health*, 6(3), e0006133.
- Kimball, A., Huang, W., Saraav, I., Funkhouser-Jones, L., Greigert, V., Yang, F., ... & Sibley, L. D. (2026). New tools for exploring parasite biology and elucidating host-pathogen interactions in cryptosporidiosis. *Microbiology and Molecular Biology Reviews*, e00148-22.
- Kumar, M. (2025). Immunocompromised Patients. *Current Topics in Emerging and Reemerging Zoonoses*, 105.
- Kundu, R., Bansal, Y., & Singla, N. (2024). The zoonotic potential of fungal pathogens: another dimension of the one health approach. *Diagnostics*, 14(18), 2050.
- Le Hingrat, Q., Sereti, I., Landay, A. L., Pandrea, I., & Apetrei, C. (2021). The Hitchhiker's Guide to CD4+ T-Cell Depletion in lentiviral infection. A critical review of the dynamics of the CD4+ T cells in SIV and HIV infection. *Frontiers in Immunology*, 12, 695674.
- Lionakis, M. S., Drummond, R. A., & Hohl, T. M. (2023). Immune responses to human fungal pathogens and therapeutic prospects. *Nature Reviews Immunology*, 23(7), 433-452.
- Ly, T., Cao, W., & Li, T. (2021). HIV-related immune activation and inflammation: current understanding and strategies. *Journal of Immunology Research*, 2021(1), 7316456.
- McCune, J. M. (2001). The dynamics of CD4+ T-cell depletion in HIV disease. *Nature*, 410(6831), 974-979.
- Mishra, S., Dwivedi, S. P., Dwivedi, N., & Singh, R. B. (2009). Immune response and possible causes of CD4+ T-cell depletion in human immunodeficiency virus (HIV)-1 infection. *The Open Nutraceuticals Journal*.
- Miti, S., Handema, R., Mulenga, L., Mwansa, J. K., Abrams, E., Frimpong, C., ... & Denison, J. A. (2020). Prevalence and characteristics of HIV drug resistance among antiretroviral treatment (ART) experienced adolescents and young adults living with HIV in Ndola, Zambia. *PLoS One*, 15(8), e0236156.
- Mu, W., Patankar, V., Kitchen, S., & Zhen, A. (2024). Examining chronic inflammation, immune metabolism, and T cell dysfunction in HIV infection. *Viruses*, 16(2), 219.
- Mubareka, S., Amuasi, J., Banerjee, A., Carabin, H., Copper Jack, J., Jardine, C., ... & Jane Parmley, E. (2023). Strengthening a One Health approach to emerging zoonoses. *Facets*, 8(1), 1-64.
- Mukherjee, K., & Mishra, D. (2026). Infection Susceptibility and Clinical Burden in Patients with Acquired Immune Dysfunction. *Archives of Medical Reports*, 3(2), 1-9.
- Navasardyan, I., Abdou, A., Kades, S., Misakyan, Y., Ochsner, J., Subbian, S., & Venketaraman, V. (2023). Tuberculosis meningitis coexisting with HIV infection: a comprehensive review. *Frontiers in Tuberculosis*, 1, 1242869.
- Obeagu, E. I., & Obeagu, G. U. (2024). Implications of B lymphocyte dysfunction in HIV/AIDS. *Elite Journal of Immunology*, 2(1), 34-46.
- Obeagu, E. I., & Obeagu, G. U. (2026). CD4/CD8 ratios as immune architects—building defenses against HIV: A narrative review. *Medicine*, 105(5), e47396.
- Okoye, A. A., & Picker, L. J. (2013). CD 4+ T-cell depletion in HIV infection: mechanisms of immunological failure. *Immunological Reviews*, 254(1), 54-64.
- Ortiz-Millán, G. (2025). One Health in a globalized world: challenges and responses to zoonotic threats. *Global Bioethics*, 36(1),

2550805.

- Overgaauw, P. A., Vinke, C. M., Van Hagen, M. A., & Lipman, L. J. (2020). A one health perspective on the human–companion animal relationship with emphasis on zoonotic aspects. *International Journal of Environmental Research and Public Health*, 17(11), 3789.
- Peng, X., Ouyang, J., Isnard, S., Lin, J., Fombuena, B., Zhu, B., & Routy, J. P. (2020). Sharing CD4+ T cell loss: when COVID-19 and HIV collide in the immune system. *Frontiers in Immunology*, 11, 596631.
- Rauf, U., Fakhar, K., Rafique, N., Mehnaz, S., Yasin, A., Fatima, T., & Niazi, S. Z. K. (2023). *Fungal zoonosis and one health*. Zoonosis, Unique Scientific Publishers, Faisalabad, Pakistan, 4, 407-419.
- Russotto, Y., Micali, C., Pellicano, G. F., Nunnari, G., & Venanzi Rullo, E. (2022). HIV and Mediterranean zoonoses: a review of the literature. *Infectious Disease Reports*, 14(5), 694-709.
- Saka, R., Domfeh, E. A., Hayford, F. E., Asante, M., Amoah, A. G., Asare, G. A., & Wiredu, E. K. (2023). Nutritional status and effect of highly active anti-retroviral therapy (HAART) on selected trace elements in people living with HIV in Ghana. *Scientific African*, 19, e01586.
- Samadi, A., & Hailat, N. (2021). Zoonotic diseases: a One Health perspective. *CABI Reviews*, (2021).
- Scaglioni, S. (2025). *The role of CD4+ T memory subsets in HIV infection: immune response in people with HIV and latency establishment*.
- Silva, D. L., Peres, N. T., & Santos, D. A. (2025). Key fungal coinfections: epidemiology, mechanisms of pathogenesis, and beyond. *MBio*, 16(5), e00562-25.
- Stump, D. S., & Vande Woude, S. (2007). Animal models for HIV AIDS: a comparative review. *Comparative Medicine*, 57(1), 33-43.
- Su, B., Kong, D., Yang, X., Zhang, T., & Kuang, Y. Q. (2022). Mucosal-associated invariant T cells: a cryptic coordinator in HIV-infected immune reconstitution. *Journal of Medical Virology*, 94(7), 3043-3053.
- Sun, Y., & Wang, L. (2024). Development of anti-HIV therapeutics: from conventional drug discovery to cutting-edge technology. *Pharmaceuticals*, 17(7), 887.
- Teer, E., Mukonowenzou, N. C., & Essop, M. F. (2025). HIV, inflammation, and immunometabolism: A model of the inflammatory theory of disease. *Viruses*, 17(6), 839.
- Tokarev, A., Machmach, K., Creegan, M., Kim, D., Eller, M. A., & Bolton, D. L. (2022). Single-Cell Profiling of Latently SIV-Infected CD4+ T Cells Directly Ex Vivo to Reveal Host Factors Supporting Reservoir Persistence. *Microbiology Spectrum*, 10(3), e00604-22.
- Tolomeo, M., & Cascio, A. (2024). The complex dysregulations of CD4 T cell subtypes in HIV infection. *International Journal of Molecular Sciences*, 25(14), 7512.
- Ullah, M. F. (2026). Anthropogenic influence, microbes and zoonotic diseases: Ecological imbalance, diverse impact and the One Health approach. *Journal of Vector Borne Diseases*, 63(1), 1-15.
- Vidya Vijayan, K. K., Karthigeyan, K. P., Tripathi, S. P., & Hanna, L. E. (2017). Pathophysiology of CD4+ T-cell depletion in HIV-1 and HIV-2 infections. *Frontiers in Immunology*, 8, 580.
- Wang, Z., Yin, X., Ma, M., Ge, H., Lang, B., Sun, H., ... & Jiang, Y. (2021). IP-10 promotes latent HIV infection in resting memory CD4+ T cells via the LIMK-cofilin pathway. *Frontiers in Immunology*, 12, 656663.
- Wilcox, B. A., & Steele, J. A. (2021). *One Health and emerging zoonotic diseases: Framework, integration and challenges*. In Handbook of global health (pp. 1-49). Cham: Springer International Publishing.
- Yan, L., Xu, K., Xiao, Q., Tuo, L., Luo, T., Wang, S., ... & Yang, X. (2023). Cellular and molecular insights into incomplete immune recovery in HIV/AIDS patients. *Frontiers in Immunology*, 14, 1152951.
- Yang, X., Su, B., Zhang, X., Liu, Y., Wu, H., & Zhang, T. (2020). Incomplete immune reconstitution in HIV/AIDS patients on antiretroviral therapy: Challenges of immunological non-responders. *Journal of Leukocyte Biology*, 107(4), 597-612.

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